

**Fig. 1 – 5: Pseudoheliozoa.** Pseudoheliozoa are a poorly defined group of minute organisms with characteristics similar to those of the Heliozoa. The main difference is the lack of axopodia with microtubular axonemes. The pseudoheliozoans have filose pseudopodia and a stalked test. **1:** The rare *Hedriocystis brachypous* is about 11  $\mu$ m across and has a polygonal test with a short stalk containing a fine filament (FI). The filopodia emerge from small perforations in the test (arrow). **2:** *Hedriocystis pellucida* is about 15  $\mu$ m across and is quite similar to *H. brachypous*, but has a long stalk. **3 - 5:** *Clathrulina elegans* is about 30  $\mu$ m across and lives in a stalked test with polygonal pores, from which the filopodia emerge. The specimen shown is in a late stage of division (5). FI – filament, N – nucleus, ST – stalk.





Fig. 1 – 3: Pseudoheliozoa. Nuclearia caulescens has many radiating, filose pseudopodia (1, arrows). The globular cell is attached to the substrate by a long, hollow stalk (1). The filopodia are granulated by minute extrusomes (1), like in heliozoans. The cells are 20–40 µm across and are covered by a 10-20 µm thick mucous layer (2, arrowheads). The irregular outline of this species is caused by amoeboid shape changes and several contractile vacuoles. Nuclearia caulescens feeds on flagellates and small ciliates (3, arrow). CV - contractile vacuoles, EX - extrusomes, N - nucleus, ST - stalk.



**Fig. 1 – 6: Ciliates.** Loxodes rostrum (1, 2, 4) and L. striatus (3, 5, 6) are middlesized (length ~ 200 µm) karyorelictean ciliates, that is, the macronucleus does not divide but originates from a micronucleus anlage. The two species differ mainly by the nuclear apparatus: two macronucleus nodules with one micronucleus in between in L. rostrum (2, 4) and two widely separate macronucleus nodules each with a micronucleus in L. striatus (5). Arrows in figure 6 mark the Müller organelles, that is, barium globules in a vacuole, which are likely gravity receptors. A – symbiotic algae, MA – macronucleus nodules, MI – micronucleus, OA – oral apparatus.







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µm long suctorian ciliate, *Sphaerophrya* stentori. AM – adoral zone of membranelles, MA – macronucleus.





**Fig. 1 – 6: Ciliates.** *Spirostomum* spp. are highly contractile, heterotrichous ciliates with a long adoral zone of membranelles (2, 3, arrows). Species are distinguished by body size and shape and the nuclear pattern. **1:** *Spirostomum ambiguum* reaches 2 mm and has a moniliform macronucleus. **2, 5:** *Spirostomum teres* is up to 400  $\mu$ m long and has an ellipsoidal macronucleus (5). **3:** *Spirostomum minus* is up to 600  $\mu$ m long and has a moniliform macronucleus. **4:** *Spirostomum semivirescens* is about 1 mm and has symbiotic algae. **6:** *Spirostomum caudatum* reaches a length of 200–300  $\mu$ m long and has a long tail (arrowheads, inset), well recognizable in this freely motile specimens. AM – adoral membranelle, CV – contractile vacuole with long canal, MA – macronucleus.



the microaerobic Sphagnum mud of Simmelried. It is related to Blepharisma and Spirostomum, as indicated by the long adoral zone of membranelles and the ability to contract (2, 5). The green (1) or pink (3) colour is caused by symbiotic algae (1) and ingested rhodobacteria (3). Occasionally, the macronucleus is parasitized and inflated (4, arrow). A - symbiotic algae, AM - adoral zone of membranelles, CV - contractile vacuole, UM - undulating membrane.





**Fig. 1 – 5: Ciliates.** Euplotes patella (1, 3 – 5) and *E. diadalos* (2), which belong to the hypotrichs, are about 100 µm long, strongly flattened (5) and differ from each other by the symbiotic green algae. *Euplotes* has an armour-like cortex and cirri on the ventral side (3), while rows of minute bristles cover the dorsal side (4). AM – adoral zone of membranelles, CC – caudal cirri, CV – contractile vacuole, TC – transverse cirri.





**Fig. 1 – 5: Ciliates.** Holosticha pullaster (1, 2) and Paraurostyla weissei (3 – 5) are hypotrichs with the cilia bundled to cirri. **1, 2:** Holosticha pullaster is about 80 µm long and easily identified by the subterminal location of the contractile vacuole (arrow) and the zigzaging midventral cirri (arrowhead). **3 – 5:** Paraurostyla weissei is 200–250 µm long and yellowish due to minute cortical granules, likely mucocysts (4). Details of the cirral pattern and oral apparatus are well recognizable in the scanning electron microscope. AM – adoral zone of membranelles, FC – frontal cirri, LMR, RMR – left and right row of marginal cirri, MA – macronucleus nodules, TV – transverse cirri, UM – undulating membrane, V1–5 – ventral cirral rows.



**Fig. 1 – 4: Ciliates.** *Pelagostrombidium viridis* is an about 80 µm long planktonic, oligotrichous ciliate, which is greenish to brownish due to cleptoplasts (stolen chloroplasts from prey algae) still working within the predator. The mighty adoral zone of membranelles (AM), which drives the organism and collects food, is well recognizable in the scanning electron microscope (2, 4). Arrowhead in figure 4 marks mouth entrance. AM – adoral zone of membranelles, TR – trichocysts.





**Fig. 1 – 5: Ciliates.** *Metopus* and its relatives have a high diversity (see also next plates). They are anaerobics living in the organic mud, especially in ponds with decaying leaves and in the anaerobic sediment of lakes. **1, 2:** *Metopus propagatus* is a massive, 150–200 µm long, tailed species. **3:** *Metopus vestitus* is a delicate, 70–80 µm long species with a caudal spine. **4:** Conjugation (sex) of *M. pulcher.* **5:** *Tropidoatractus acuminatus* is about 120 µm long and distinctly spiralized. AM – adoral zone of membranelles, CV – contractile vacuole, MA – macronucleus.



**Fig. 1 – 4: Ciliates.** Diversity of metopids, obligate anaerobics in the mud of ponds. **1, 3:** *Metopus bacillatus* (or *M. striatus*; courtesy Prof. C. F. Bardele), ventral (1) and dorsal (3) view in the SEM and interference contrast, where the bacilli-shaped mucocysts appear as minute, bright dots (3). SEM shows the real beauty of this type of ciliates. **2:** *Metopus ovalis* is about 150 µm long and a typical member of the genus (courtesy Prof. Bardele). **4:** *Luido parvulus*, an about 40 µm long, aberrant metopid, has a very short adoral zone (arrowheads) and long cilia bundles. Inset: varied focal plane. AM – adoral zone of membranelles, MA – macronucleus, PZ – perizonal ciliary array, UM – undulating membrane.



**Fig. 1 – 6: Ciliates.** Further metopids (see foregoing plates). **1, 2:** *Metopus nasutus* is about 130 μm long and has a nose-like process (arrow). **3:** *Metopus laminarius* is up to 260 μm long and one of the largest members of the genus. The adorale zone of membranelles is distinctly spiralized. The shape of the cells can be irregular by masses of feeded rhodobacteria. **4:** *Metopus acidiferus* is about 100 μm long and a typical member of the group. **5:** *Metopus mucicola* is a tiny (~ 40 μm) species with a hyaline, slimy lorica. **6:** *Metopus campanula* is about 60 μm long. AM – adoral zone of membranelles, CC – caudal cilia, CV – contractile vacuole, FV – food vacuoles, MA – macronucleus.



**Fig. 1 – 4: Ciliates.** The "aliens" shown on this plate belong to the odontostomatids, a highly aberrant group living only in anaerobic (sapropelic) mud. **1:** *Caenomorpha medusula*, length 150 μm. **2:** *Caenomorpha sapropelica*, length 200 μm. **3:** *Caenomorpha uniserialis*, posterior polar view in the scanning electron microscope, long axis about 60 μm. Arrowheads mark begin and end of adoral zone of membranelles. **4:** *Discomorphella pectinata* has curious ventral spines (arrowheads) and a conspicuous ridge with short ciliary rows (arrow), length 80 μm. CV – contractile vacuole, FV – food vacuole with rhodobacteria, MA – macronucleus nodules.



**Fig. 1 – 5: Ciliates.** This plate shows some odontostomatids, which have a highly sculptured cortex and complex ciliature (see also foregoing plate). **1, 2:** *Epalxella antiquorum*, length about 80 µm, right (2) and left (1) side view showing the three typical concavities (arrowheads). **3:** *Mylestoma discoideum* has a discoidal body about 40 µm across. **4:** *Mylestoma anatinum* is about 25 µm long and has a ventral anterior beak. **5:** *Mylestoma pusillum* is a truely minute ciliate (~ 20 µm) with a conspicuous ciliary bundle in body midline. CV – contractile vacuole, MA – macronucleus, OA – site of oral apparatus.



**Fig. 1 – 6: Ciliates.** Odontostomatids (see also previous plates) are obligate anaerobics in the mud of ponds and lakes. They have complex body shapes and ciliary patterns. **1, 2, 4:** *Saprodinium dentatum* is about 70 μm long and has 8 (numerals) cortical processes posteriorly. **3:** Right side view of *Pelodinium reniforme*, which is about 50 μm long and has a highly characteristic crest (ciliary) pattern. **5, 6:** *Atopodinium fibulatum* is only 40 μm long and has a conspicuous posterior indentation when viewed ventrally (6). CC – caudal cilia, OA – site of oral apparatus.





Fig. 1 – 4: Ciliates. Phacodinium metchnikoffi is a rare, about 150 µm long moss ciliate of doubtful phylogenetic affinity. Possibly, it is related to the hypotrichs (Stylonychia, Euplotes) because the left side has bristle rows (2) and the cilia are grouped to cirri (3, 4). The surface is strongly ridged, especially on the left (dorsal) side (2). 1, 4: Right side views, showing the adoral zone of membranelles (AM) extending almost whole body length. 2: Posterior polar view. 3: Detail of a ciliary row. AM - adoral zone of membranelles, BR - bristle rows, CV - contractile vacuole, FV - food vacuole.







Fig. 1 – 5: Ciliates. Epispathidium amphoriforme (1 - 3) and Apertospathula armata (4, 5) belong to the holotrichously ciliated spathidiids. The anterior body end is modified to an oral apparatus, whose bulge (OB) contains toxicysts (EX), used to capture protists and small metazoans. 1 - 3: Epispathidium amphoriforme is about 150 µm long and has an inconspicuous dorsal brush (B), that is, three modified ciliary rows with short, paired cilia in the anterior region of the dorsal side. The function of this organelle is not known. Further details, see next plate. 4, 5: The tiny A. armata is only 60 µm long and the short oral bulge contains minute ( $\sim 1 \mu m$ ) toxicysts (5, EX). B - dorsal brush, CV - contractile vacuole, MA - macronucleus, OB - oral bulge.



**Fig. 1, 2: Ciliates.** *Epispathidium amphoriforme*, ciliary pattern after protargol silver impregnation (see also foregoing plate). Silver impregnation reveals the basal bodies of the cilia with perfect clarity and is thus the most important tool in ciliate alpha-taxonomy. The micrographs show the left and right mouth area, that is, the circumoral ciliary row (CK) and the somatic ciliary rows, which are strongly curved and densely ciliated anteriorly. The paired basal bodies of the dorsal brush (B) are well recognizable. B – dorsal brush, CK – circumoral ciliary row (kinety), OB – oral bulge.



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about 200  $\mu$ m long and green due to symbiotic algae. The mouth contains many rod-shaped toxicysts (5). CV – contractile vacuole, MA - macronucleus, ÓB - oral bulge.



**Fig. 1 – 3: Ciliates.** *Homalozoon vermiculare* is a highly contractile (2, 3), gymnostome ciliate, which can extend to 600 µm (1). The ciliature of the left side is reduced to three rows of minute bristles (1, 3, arrowheads). The hemispherical oral bulge contains many toxicysts to capture other ciliates (1, inset, arrows). The function of the pharyngeal mass (1, arrows) is not known. CV – contractile vacuole, EX – extrusomes, MA – moniliform macronucleus, OB – oral bulge.





**Fig. 1 – 5: Ciliates. 1, 2:** *Trachelius ovum* is an about 250 µm long, gymnostome ciliate, which feeds on colonial peritrichs. It is related to *Dileptus* and thus has a short proboscis, on which the oral bulge (OB) ends at the oral opening (2, OO). In mid-body, there is a more densely ciliated groove (1, asterisk), the function of which is not known. **3:** *Prorodon niveus* is an about 350 µm long, laterally flattened, gymnostome ciliate with long extrusomes in the mouth. **4, 5:** *Condylostomides tardus* is a colourless, about 200 µm long heterotrichous ciliate with three macronucleus nodules (5). AM – adoral zone of membranelles, CV – contractile vacuole, EX – extrusomes, FV – food vacuole, MA – mmacronucleus, OA – oral apparatus, OB – oral bulge, OO – oral opening.









**Fig. 1 – 4: Ciliates.** *Kreutzophrya sphagnicola* is a new genus and species discovered in the mud of Simmelried. It is 200–250 µm long (3), holotrichously ciliated (1), and has a small, oblique mouth (1 – 3) with rod-shaped toxicysts (4, T). Mouth details, not recognizable on the micrographs, define the new genus *Kreutzophrya* dedicated to M. Kreutz by W. Foissner.

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**Fig. 1 – 7: Ciliates.** These are rapacious, about 100  $\mu$ m long, holotrichously ciliated, rare haptorids. **1 – 3:** *Actinobolina vorax* (1) and *Dactylochlamys pisciformis* (2, 3) have retractile tentacles each containing a toxicyst (arrowheads). The ciliary rows extend obliquely (3). **4 – 7:** Two, likely undescribed *Legendrea* species. They have warts (4, 5, arrowheads) with short extrusomes, while the oral bulge, which spirals around the cell (asterisks), contains long extrusomes (7, arrows). CV – contractile vacuole, MA – macronucleus, OB – oral bulge.





**Fig. 1 – 6: Ciliates.** Undescribed gymnostomes (haptorids). **1:** A broad *Penardiella* with distinct, but hardly flattened extrusome fringe (EX), length 110  $\mu$ m. **2, 3:** Right side and ventral view of an *Arcuospathidium* or *Apertospathula*, length 190  $\mu$ m. **4:** A curious species likely representing a new genus, length 100  $\mu$ m. **5:** A tiny, new *Perispira*, length 60  $\mu$ m. **6:** A small *Spathidium* or *Arcuospathidium*, length 60  $\mu$ m. CV – contractile vacuole, EX – extrusomes, MA – macronucleus, OB – oral bulge.



**Fig. 1 – 7: Ciliates.** Undescribed species. **1, 2:** A curiously-shaped, large pleurostome haptorid, length 240  $\mu$ m. **3:** A warty spathidiid, length 75  $\mu$ m. **4, 7:** A nice *Perispira* or *Bryophyllum*-like haptorid with conspicuous dorsal brush (arrowheads), length 160  $\mu$ m. **5:** A prostomatid with many very narrowly spaced ciliary rows, length 130  $\mu$ m. **6:** A globular haptorid with large oral bulge, length 40  $\mu$ m. EX – extrusomes, MA – macronucleus, MI – micronucleus, OB – oral bulge.

![](_page_33_Figure_0.jpeg)

**Fig. 1 – 5: Ciliates.** Amphileptus procerus is a pleurostome ciliate with a laterally located mouth slit (1) and a fully ciliated right side, while the left has rows of minute bristles (3 – 5). Arrow in (5) marks region shown at higher magnification in figure 3. *Amphileptus procerus* is up to 500 µm long and highly contractile as shown by figures 1 (contracted) and 2 (fully extended). BR – bristles, DB – dorsal brush.

![](_page_34_Picture_0.jpeg)

**Fig. 1, 2: Ciliates.** *Loxophyllum meleagris*, an about 400 µm long, majesticly gliding, pleurostomatid ciliate, is easily recognizable by the large size, the moniliform macronucleus, the curious toxicyst warts (for a detail see next plate), and the naked left side, which contains rows of minute bristles. The arrow marks the proximal end of the mouth, while the arrowheads denote the contractile vacuole which has a canal extending anteriorly. BR – bristle rows, CV – contractile vacuole, DB – dorsal brush, MA – moniliform macronucleus, W – extrusome warts.

![](_page_35_Picture_1.jpeg)

![](_page_35_Picture_2.jpeg)

Fig. 1 – 3: Ciliates (read in context with preceding plate). 1, 2: Loxophyllum helus is smaller (length ~ 200 µm) than L. meleagris and has only two macronucleus nodules. The extrusome warts are as distinct as in *L. meleagris* shown in figure 2. This SEM micrograph shows also a typical feature of the pleurostomatid ciliates: only the right side is ciliated, while the left has rows of minute ciliary stumps. Arrows mark excretory pores of the contractile vacuole. 3: Opisthodon niemeccense is a rare, about 150 µm long pleurostome with an elliptical shape and a small anterior groove (arrowhead) containing the dorsal brush. The arrow marks the serrate body margin, a prominent feature not described in the literature; thus the Simmelried population might be a distinct species. BR - bristle rows, MA - macronucleus, W - extrusome warts.

![](_page_36_Picture_0.jpeg)

**Fig. 1 – 4: Ciliates.** *Trithigmostoma cucullus* from life (4) and in the scanning electron microscope. *Trithigmostoma* is a cyrtophorid, about 100 µm long ciliate, which feeds on diatoms (4) ingested via the oral basket (cyrtos; BA in 3, 4). As an Aufwuchs dweller, it is ciliated mainly on the ventral side (1), the dorsal bears only a short (sensory?) ciliary row (2; arrow). BA – oral basket, CV – contractile vacuoles.

![](_page_37_Picture_0.jpeg)

**Fig. 1 – 4: Ciliates.** *Pseudochilodonopsis piscatoris* (1; ventral view, length ~ 70  $\mu$ m) and *Chilodonella uncinata* (2 – 4; ventral and dorsal views, length ~ 40  $\mu$ m) are cyrtophorid ciliates, that is, have a conspicuous oral basket (BA). *Chilodonella uncinata* is the classical ciliate, where B. M. Klein discovered the silverline pattern revealed with silver nitrate (3, 4). B – dorsal brush, BA – oral basket, FV – food vacuole, MA – macronucleus, arrowheads – contractile vacuoles.

![](_page_38_Picture_0.jpeg)

**Fig. 1 – 7: Ciliates.** Although the adult suctoria lack any cilia, they are true ciliates because they have a ciliated swarmer and the typical ciliate nuclear apparatus composed of a macronucleus and a micronucleus (2). The adult suctoria have numerous tentacles with extrusomes (haptocysts) at the distal end to catch the prey. **1 – 7:** *Metacineta cuspidata* is 30–40 µm high and rare in Simmelried. The lorica has a tapered stalk (1) and four apical clefts (3, arrows), where bundles of tentacles emerge. The distal region of the tentacles is bent or curled in a characteristic, species-specific manner (4, arrowheads; inset). The sole-shaped swarmer is 30–35 µm long and dorsoventrally flattened. The ventral side (5) is ciliated, while the dorsal side and the posterior end bear some small tentacles (6, TC) and long cilia (6, arrowheads). The new stalk and lorica will be generated by the scopuloid when the swarmer develops to the adult (7, SC). CV – contractile vacuole, MA – macronucleus, MI – micronucleus, SC – scopuloid, TC – tentacles.

![](_page_39_Picture_0.jpeg)

**Fig. 1 – 7: Ciliates. 1, 2:** *Metacineta mystacina* var. *brevipes* is 30 µm across and can be distinguished from *M. cuspidata* (see previous plate) by the straight tentacles (1, arrowheads) and the six apical clefts (2, arrows). The lorica has a short, tapered stalk (1, inset) and measures  $40-45 \ \mu m$ . **3 – 5:** *Parapodophrya soliformis* is  $40-100 \ \mu m$  across and the most common suctor in Simmelried. It has a spherical body with radiating tentacles. Retracted tentacles appear as tapered cones 2–3 µm high (4, arrowheads). The prey (in this case the ciliate *Holophrya* sp.) can be much larger than the predator and is sucked by the tentacles (5, arrows). **6, 7:** *Heliophrya minima* (20–50 µm) is a discoidal, strongly flattened suctor attached to the substrate with the whole body. The tentacles are separate and emerge from the margin (6). CV – contractile vacuole, MA – macronucleus.

![](_page_40_Picture_0.jpeg)

**Fig. 1 – 6: Ciliates.** Some suctoria parasitize other ciliates. They penetrate the host via the mouth or produce a deep host invagination connected to the environment by a minute tube. After feeding, they divide and generate swarmers. **1 – 4:** Some stages of the life cycle of *Sphaerophrya parurolepti*, which parasitizes *Uroleptus caudatus*, a stichotrichous ciliate. **1:** An about 12 µm-sized adult (arrow) attached to the mouth area. **2:** A squashed host with a 27 µm-sized parasite. **3:** A squashed host with four parasites. **4:** Three parasites leaving the host. **5, 6:** *Paramecium bursaria* parasitized by *Podophrya* or *Sphaerophrya* sp. Figure 5 shows three parasites in a bottle-shaped invagination of the host (arrow). When squashed, the globular parasite (PA) and the ellipsoidal host's macronucleus become visible (6). The micronucleus is parasitized by bacteria and thus much larger than usual. CV – contractile vacuole, EX – extrusomes, MA – macronucleus, MI – micronucleus, PA – parasite.

![](_page_41_Picture_0.jpeg)

**Fig. 1 – 7: Ciliates.** This plate shows the life cycle of *Nassulopsis elegans*, an about 200 µm long, nassulid ciliate (1, 2) feeding on filamentous cyanobacteria, which are taken (6) and transported (4, 5) into the cell by the conspicuous oral basket made of microtubular rods (6). The coloured droplets are digestion products of the cyanobacteria. After feeding, specimens conjugate (3) or encyst (7). BA – oral basket.

![](_page_42_Picture_0.jpeg)

![](_page_43_Picture_0.jpeg)

**Fig. 1 – 6: Ciliates.** Drepanomonas spp. are small, microthoracid ciliates with a nicely sculptured surface and trichocysts well recognizable in the scanning electron microscope (5, 6). **1:** Drepanomonas dentatum (70–80 µm), left side view. **2, 3:** Drepanomonas spec. (50–60 µm), a likely undescribed species from Simmelried, in left side (2) and ventral (3) view, showing the strongly sculptured surface. **4, 6:** Drepanomonas revoluta, right side and ventral view, length about 30 µm. **5:** Microthoracid ciliates, here *Pseudomicrothorax*, have curious trichocysts with four anchor-like processes. OA – entrance to oral apparatus.

![](_page_44_Picture_0.jpeg)

**Fig. 1 – 7: Ciliates.** *Microthorax* species are 20–40 µm long and strongly flattened. The glossy cortex has a complex pattern of crests and furrows, and the ciliature is strongly reduced. Most are microaerobic and feed on bacteria. **1, 2:** Right and left side view of *M. costatus* which has a curious cortex pattern (arrows). **3, 4, 6:** *Microthorax viridis* has symbiotic green algae and has a deep furrow on the anterior left side (4, arrowhead). Note the cordiform entrance to the buccal cavity (6). **5, 7:** *Microthorax pusillus* is reddish due to ingested sulphur bacteria. Note the complex cortical pattern in the buccal cavity. CV – contractile vacuole, OA – oral apparatus.

![](_page_45_Picture_0.jpeg)

**Fig. 1 – 3: Ciliates.** Bryometopus sphagni (1, 3) and *B. viridis* (2) are about 120  $\mu$ m long and differ mainly by the symbiotic algae (absent vs. present). Bryometopus belongs to the colpodid ciliates and has a conspicuous, oblique mouth well recognizable both in vivo (1, 2) and the scanning electron microscope (3). Both species prefer moss habitats and feed on algae and small ciliates. AM – adoral zone of membranelles, CV – contractile vacuole, FV – food vacuoles, UM – undulating membrane.

![](_page_45_Figure_2.jpeg)

![](_page_46_Picture_0.jpeg)

**Fig. 1 – 4: Ciliates.** *Paracondylostoma setigerum chlorelligerum* from life (1, 3) and after silver carbonate impregnation (2, 4). This about 70  $\mu$ m long, colpodid ciliate was discovered in the Simmelried and is characterized by the symbiotic green algae. Note the ciliary ribbon (R) from which long, tactile bristles orginate (arrows). A – symbiotic green algae, MA – macronucleus, R – ciliary ribbon, UM – undulating membrane.

![](_page_47_Picture_0.jpeg)

**Fig. 1 – 4: Ciliates.** *Platyophrya sphagni* (1, 2) and *Thylakidium pituitosum* (3, 4) are colpodid ciliates with symbiotic algae because they live in a poor habitat, viz., in moorland puddles. **1, 2:** *Platyophrya* is a highly flexible, about 40  $\mu$ m long ciliate with a minute, subapical oral opening (arrow). It feeds on bacteria and small protists. **3, 4:** *Thylakidium* is about 80  $\mu$ m long and has a large oral funnel (ends marked by arrowheads). Thus, it can feed on various algae. The species lives in a very hyaline, slimy lorica not recognizable in this micrograph.

![](_page_48_Figure_0.jpeg)

**Fig. 1 – 3: Ciliates.** Bursaria ovata (3), which occurs in the Simmelried, from life, and *B. truncata* (1, 2), which does not occur in the Simmelried, in the scanning electron microscope. These are giantic, up to 1 mm long, colpodid ciliates differing, inter alia, by the shape of the oral tube (OT): U-like (3) vs. semicircular. **1:** Frontal view showing the oral tube entrance. **2, 3:** Ventral views. The oral tube contains a large adoral zone (AM). AM – adoral zone, OT – oral tube.

![](_page_49_Picture_0.jpeg)

**Fig. 1 – 5: Ciliates.** *Vasicola ciliata* from life (1 – 3) and after silver carbonate impregnation to reveal the highly ordered, ornamental ciliary pattern (4, 5). This about 130 µm long ciliate lives in a delicate lorica anchored to debris. *Vasicola* is a sapropelic ciliate and feeds on rhodobacteria and algae, making the food vacuoles red or green. The cortex shows a rectangular pattern (3). MA – macronucleus, OO – oral opening.