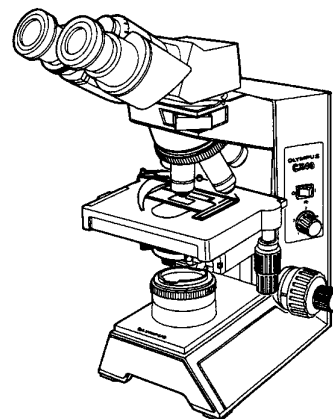


OLYMPUS®



INSTRUCTIONS

CX40

BIOLOGICAL MICROSCOPE

This instruction manual is for the Olympus Biological Microscope Model CX40. To obtain optimum performance and to familiarize yourself fully with the use of this microscope, we recommend that you study this manual thoroughly before operating the microscope. Retain this instruction manual in an easily accessible place near the work desk for future reference.



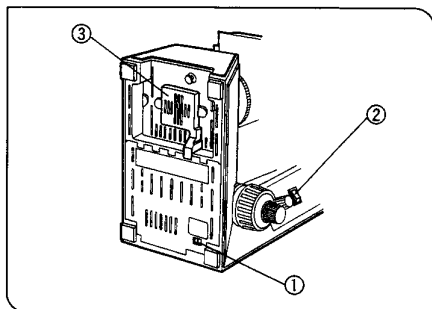
AX7018

IMPORTANT

This unit employs a UIS (universal infinity system) optical design, and should only be used with UIS eyepieces, objectives, condensers, etc. Less than optimum performance may result if inappropriate accessories are used.



SAFETY PRECAUTIONS



1. Make sure that the voltage selector switch ① located on the underside of the base of the microscope frame is set to conform with the local mains voltage. Move the switch using an Allen screwdriver, etc.
 - ⊙ When shipped from the factory, the switch is set to 100-120V and 230-240V in 100V and 200V areas, respectively.
2. To avoid potential shock hazard and fire, always switch the main switch ② to "O" (OFF) and disconnect the power cord from the AC receptacle at the rear of the microscope and from the wall outlet before replacing the bulb. Allow the lamp housing cover ③ and the bulb to cool before touching.
3. Install the microscope on a level table. The air vents on the underside of the base should never be blocked such as by placing the microscope on a flexible surface, e.g., a carpet, as this could result in overheating and cause a fire.
4. Always use the power cord provided by Olympus. If no power cord is provided, please select the proper power cord by referring to the section "PROPER SELECTION OF THE POWER SUPPLY CORD" at the end of this instruction manual. If the proper power cord is not used, Olympus can no longer warrant the electrical safety and performance of the equipment.
5. Connect the power cord correctly and ensure that the **ground terminal** of the power supply and that of the wall outlet are properly connected. If the equipment is not grounded, Olympus can no longer warrant the electrical safety and performance of the equipment.
6. Never insert metal objects, etc. into the air vents of the microscope frame as this will result in electrical shock and personal injury.

Safety symbols

The following symbols are found on the microscope. Study the meaning of the symbols, and always use the microscope in the safest possible manner.

Symbol	Explanation
	Indicates that the surface becomes hot, and should not be touched with bare hands.
	Before use, carefully read the instruction manual. Improper handling could result in injury to the user and/or damage to the equipment.
	Indicates that the main switch is ON.
	Indicates that the main switch is OFF.

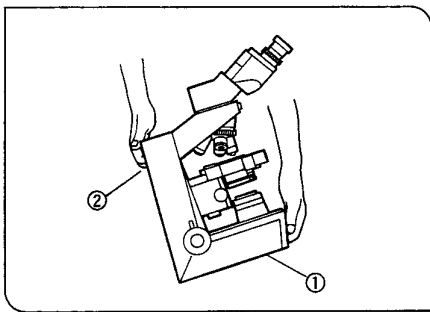
Warning Labels

Warning labels are affixed at parts where special precaution is required when handling and using the microscope. Always heed the warnings.

Warning label position	Base underside	[Caution for bulb replacement]
------------------------	----------------	--------------------------------

If warning labels become soiled, peel off, etc. contact your local Olympus representative to have them replaced.

1 Getting Ready



1. A microscope is a precision instrument. Handle it with care and avoid subjecting it to sudden or severe impact.
 2. Do not use the microscope where it is subjected to direct sunlight, high temperature and humidity, dust or vibrations. (For operating environment conditions, refer to and adhere to the conditions specified in Section 6, SPECIFICATIONS on page 17.)
 3. The tension of the coarse focus adjustment knob should only be adjusted by means of the tension adjustment ring.
 4. Heat from this microscope is led away by natural convection. Consequently, do not use it in a constricted space and ensure that the room is well ventilated.
 5. When moving the microscope, carry it with one hand under the base ① and the other hand holding at the recessed handle on the rear of the arm ② as shown in the illustration on the left.
- ★ **Damage to the microscope may occur if you hold it by the stage, binocular section of the observation tube, etc. Also make sure that eyepieces, specimen, etc. do not fall off.**
 - ★ **Sliding the microscope on the surface of the table may damage or tear off the rubber feet and/or scratch the table top surface.**

2 Maintenance and Storage

1. Clean all glass components by wiping gently with gauze. To remove fingerprints or oil smudges, wipe with gauze slightly moistened with a mixture of ether (70%) and alcohol (30%).
 - ▲ **Since solvents such as ether and alcohol are highly flammable, they must be handled carefully. Be sure to keep these chemicals away from open flames or potential sources of electrical sparks — for example, electrical equipment that is being switched on or off. Also remember to always use these chemicals only in a well-ventilated room.**
2. Do not attempt to use organic solvents to clean the non-optical components of the microscope. To clean these, use a lint-free, soft cloth lightly moistened with a diluted neutral detergent.
3. Do not disassemble any part of the microscope as malfunction or damage may occur.
4. When not using the microscope, keep it covered with the provided dust cover.

3 Caution

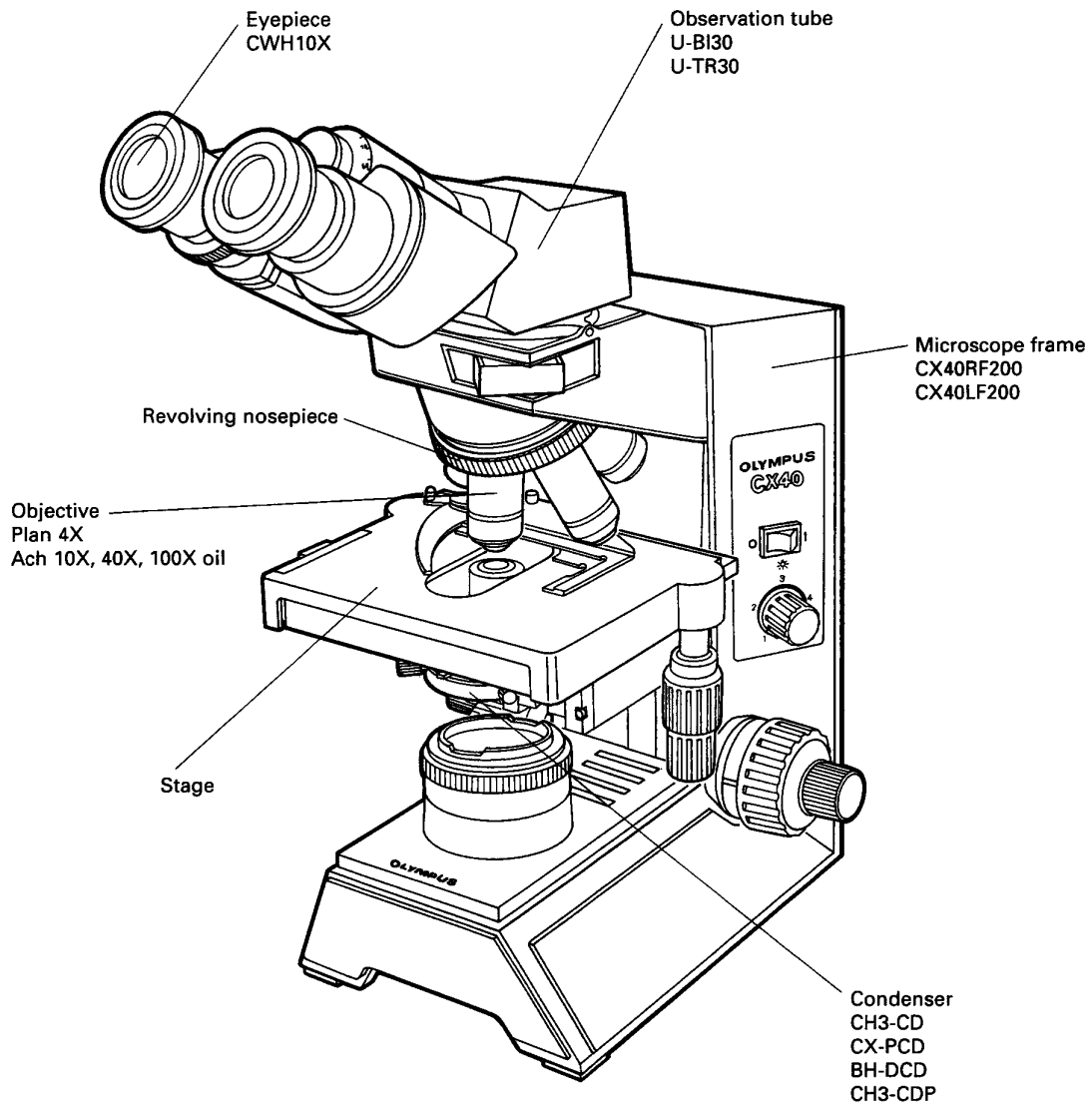
If the microscope is operated in a manner not specified by this manual, the safety of the user may be imperiled. In addition, the equipment may also be damaged. Always operate the equipment as outlined in this instruction manual.

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1

NOMENCLATURE



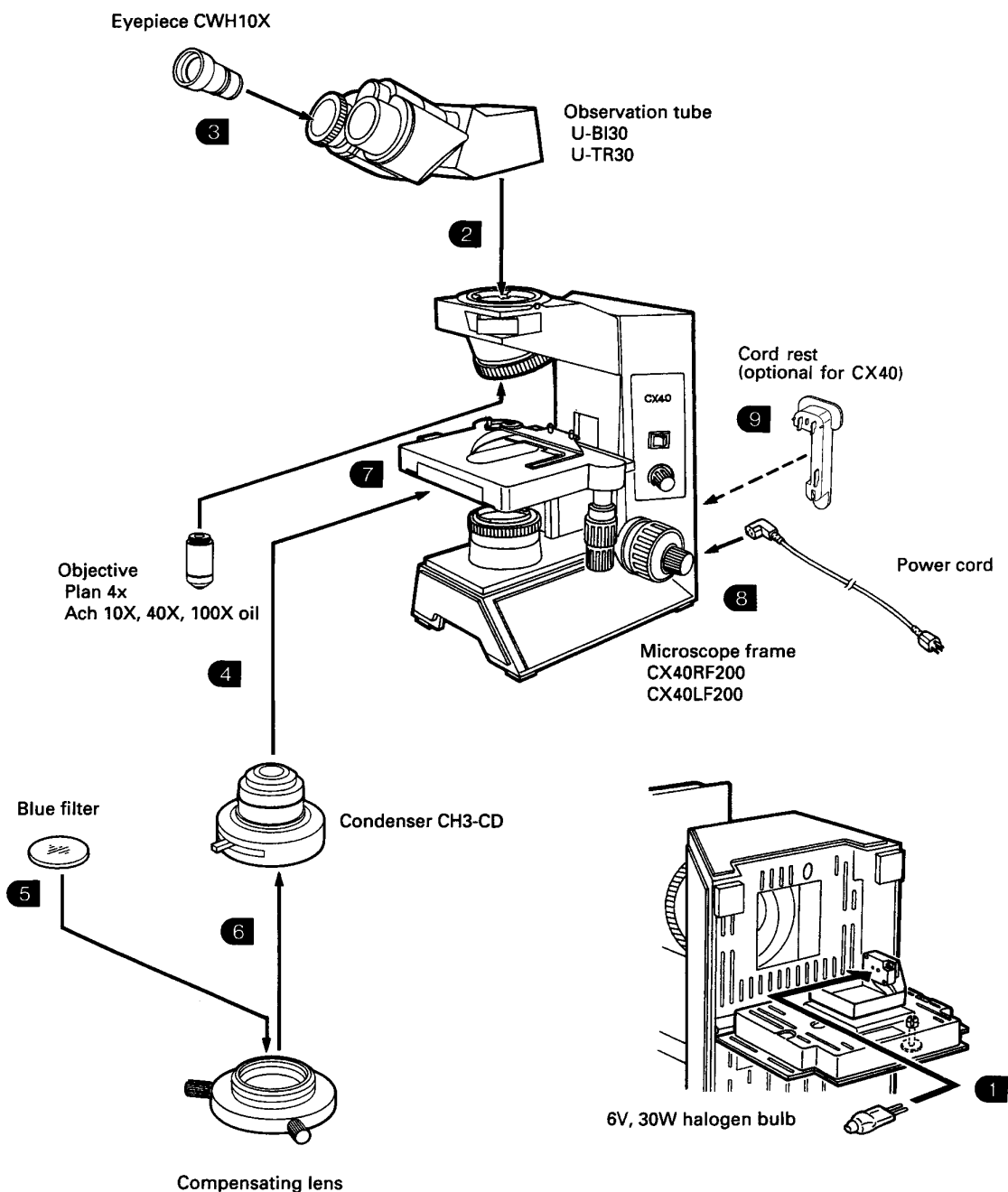
2 ASSEMBLY

2-1 Assembly Diagram

The diagram below shows how to assemble the various components. The numbers indicate the order of assembly.

★ When assembling the microscope, make sure that all parts are free of dust and dirt, and avoid scratching any parts or touching glass surfaces.

★ Retain the provided Allen screwdriver. It should be used when units are replaced or added.



2-2 Detailed Assembly Procedure

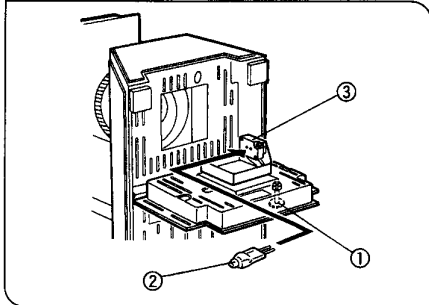


Fig. 1

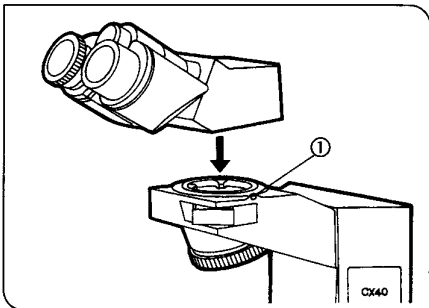


Fig. 2

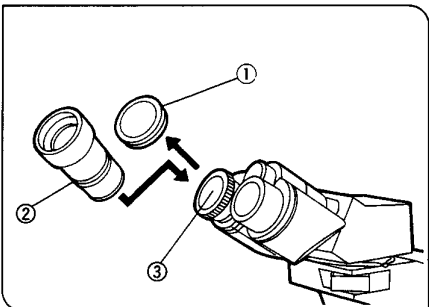


Fig. 3

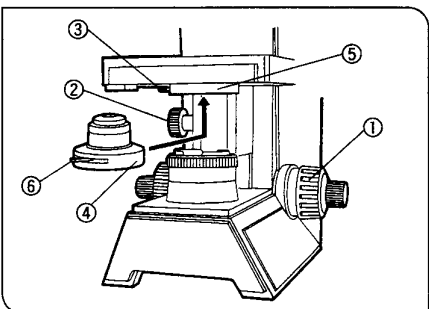


Fig. 4

1 Mounting the Bulb (Replacement of) (Fig. 1)

1. Turn the microscope frame on its side and pull the lamp housing knob ① on the underside of the base to open the lamp housing cover.
2. Holding the halogen bulb ② contained in the polyethylene bag to avoid leaving fingerprints on the bulb, fully insert the contact pins into the bulb socket ③. When properly seated, pull off the polyethylene bag.

<<Applicable bulb>>

6V, 30W halogen bulb: 6V30W HAL (Philips 5761)

⚠ Always use the designated bulb. Use of improper bulb may result in fire.

⚠ Do not touch the bulb with bare hands. If fingerprints are accidentally left on the bulb, wipe the bulb with a soft, lint-free cloth moistened with alcohol. Using a contaminated bulb will shorten the service life of the bulb.

3. With the lamp housing knob still pulled out, close the lamp housing cover. Then push in the lamp housing knob to lock the cover.

★ The lamp housing cover cannot be closed if the knob is pushed in before you attempt to close cover.

Precautions for Halogen Bulb Replacement

⚠ Whenever you replace the bulb during use or right after use, first move the main switch to "O" (OFF), disconnect the power cord from the wall outlet, and allow the bulb and parts around the bulb to cool before touching.

★ If the bulb burns out during an observation and needs to be replaced, remove eyepieces, specimen, and other objects likely to fall off, before tilting the microscope frame to replace the bulb.

2 Mounting the Observation Tube (Fig. 2)

1. Using the provided Allen screwdriver, loosen the observation tube clamping screw ①.
2. Insert the circular dovetail mount at the bottom of the observation tube into the opening on the microscope frame, positioning the observation tube to point the binocular eyepieces towards the front. Clamp the observation tube by tightening the clamping screw ①.

3 Mounting the Eyepieces (Fig. 3)

1. Remove the eyepiece dust cap ①. (Fig. 3)
2. Insert the eyepiece ② into the eyepiece sleeve ③ as far as it will go. (Fig. 3)
3. Mount the other eyepiece in the same manner.

4 Mounting the Condenser (Fig. 4)

1. Turn the coarse adjustment knob ① to raise the stage to its highest position.
2. Turn the condenser height adjustment knob ② to lower the condenser holder to the position where the clamping screw ③ can be turned.
3. Insert the condenser ④ all the way into the mounting hole ⑤ and clamp by tightening the clamping screw ③. Positioning the condenser with the diaphragm lever ⑥ at the front facilitates operation.
4. Turn the condenser height adjustment knob ② to raise the condenser holder to its highest position.

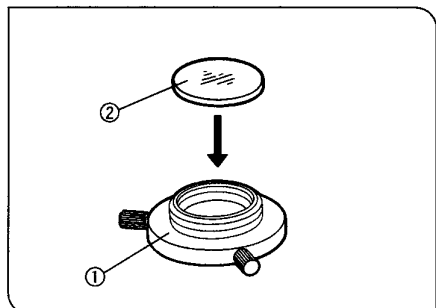


Fig. 5

5 Mounting the Blue Filter (Fig. 5)

- ⊙ The blue filter converts the color temperature of tungsten illumination into the color temperature of daylight to show the specimen in natural colors.
- Place the 32.5 mm diameter blue filter ② in the compensating lens ①.

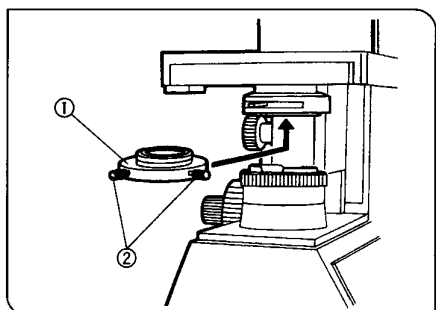


Fig. 6

6 Mounting the Compensating lens (Fig. 6)

1. Insert the compensating lens ① into the condenser from below until it seats with a click.
2. Place the lens so that the two centering screws ② are at the front.

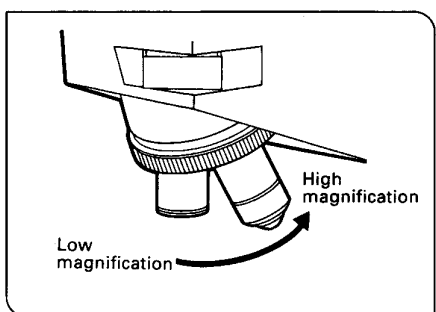


Fig. 7

7 Mounting the Objectives (Fig. 7)

- Mount the objectives on the revolving nosepiece in such a manner that the magnification increases from low to higher powers in a clockwise direction.
- ⊙ If there are empty sockets with no objective mounted, make sure to attach the provided dust plugs.

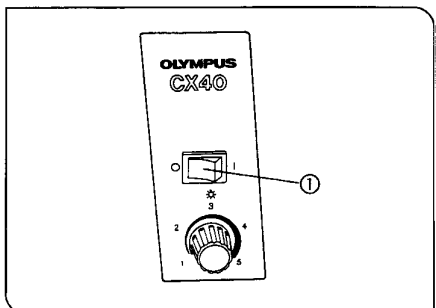


Fig. 8

8 Connecting the Power Cord (Fig. 8)

- ⚠ Cables and cords are vulnerable when bent or twisted. Never subject them to excessive force.
- ⚠ Make sure that the main switch ① is on "O" (OFF) before connecting the power cord. (Fig. 8)
- ⚠ Always use the power cord provided by Olympus. If no power cord is provided, please select the proper power cord by referring to the section "PROPER SELECTION OF THE POWER SUPPLY CORD" at the end of this instruction manual.

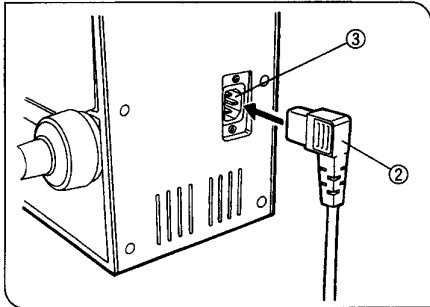


Fig. 9

1. Connect the power cord plug ② to the AC receptacle ③. (Fig. 9)

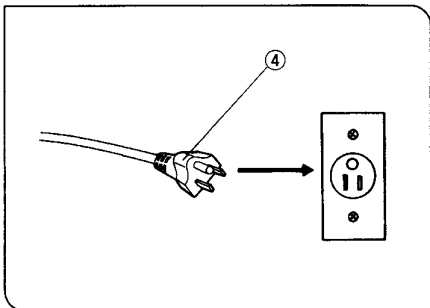


Fig. 10

2. Plug the power cord plug ④ into a wall outlet. (Fig. 10)

⚠ Connect the power cord correctly and ensure that the ground terminal of the power supply and that of the wall outlet are properly connected. If the equipment is not grounded, Olympus can no longer warrant the electrical safety and performance of the equipment.

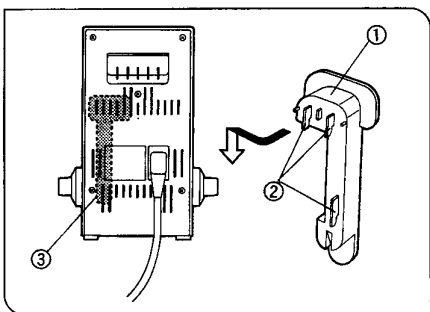


Fig. 11

9 Mounting the Condenser (Fig. 11,12)

When the cord rest (CH3-CH) is attached on the rear panel of the microscope frame, the power cord can be wound around it for storage.

Align the hooks ② on the cord rest ① with the air vent slits at ③, and insert the hooks into the slits until the cord rest is pressed against the rear panel of the microscope frame. Then pull the cord rest down to secure it.

Note: The illustration of the cord rest only shows the sections relevant for the explanation here. The actual cord rest looks somewhat different.

★ Do not grasp the cord rest when moving the microscope. The cord rest may suddenly come loose and you may drop the microscope causing material damage or personal injury.

Detaching - Move the microscope frame to the edge of the table ④. Then apply a screwdriver ⑤ or Allen wrench to the lower portion of the cord rest ① and push in the ① ② directions to move the entire cord rest upward to allow it to be detached.

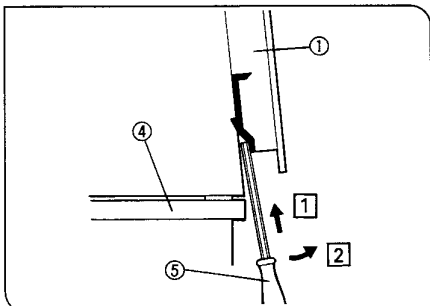
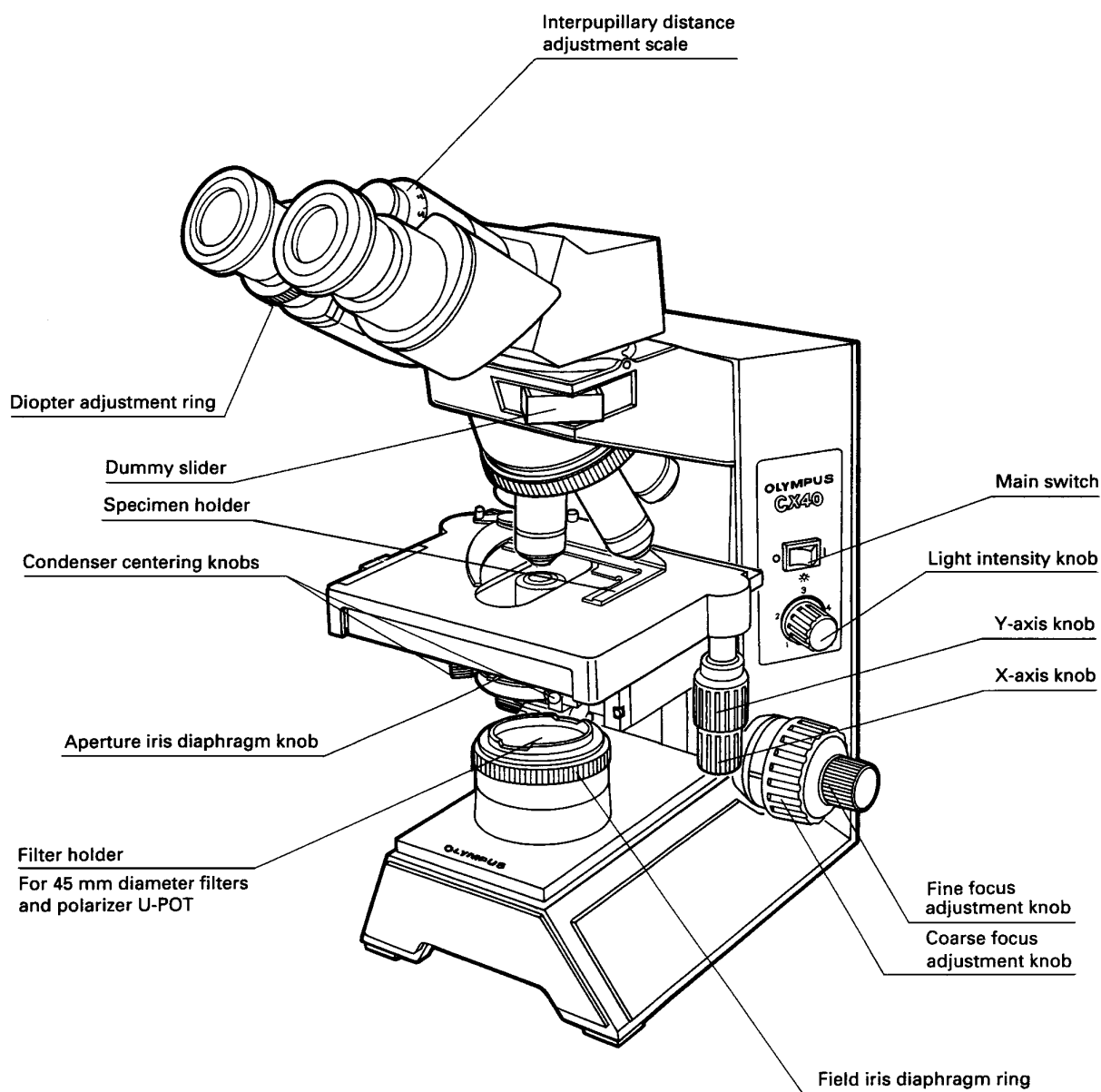


Fig. 12

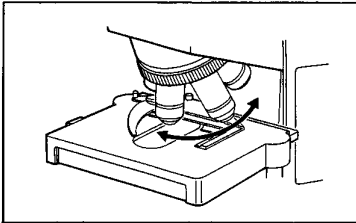
3 CONTROLS

© The explanations are based on the CX-RF200. The CX-LF200 has the stage knobs on the left side.

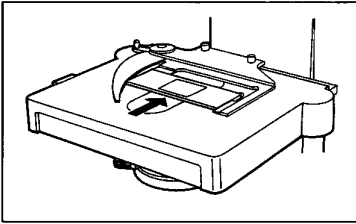


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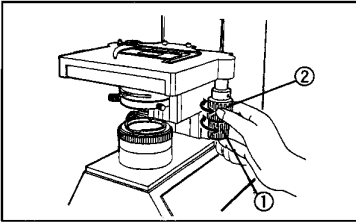
SUMMARY OF OBSERVATION PROCEDURES



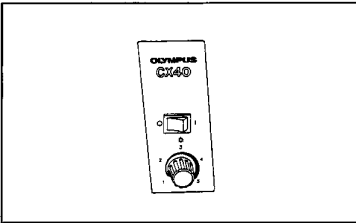
1. Turn the revolving nosepiece to engage the 10X objective. Make sure that the revolving nosepiece stops with an audible click.



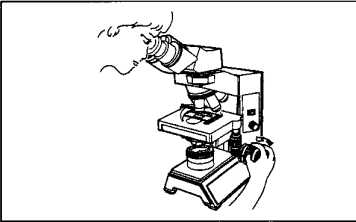
2. Place a specimen on the stage. (Page 10)



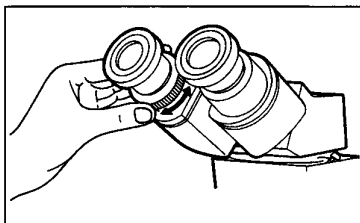
3. Turn the X-axis knob ① and Y-axis knob ② to move the specimen into the light path. (Page 10)



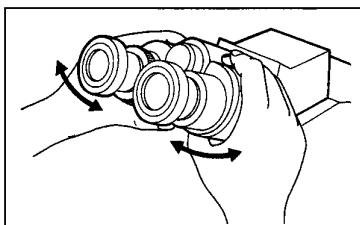
4. Switch the main switch to "I" (ON) and adjust the brightness with the light intensity knob. (Page 9)



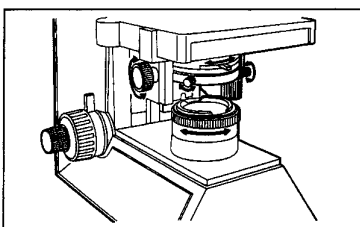
5. Looking through the right eyepiece with your right eye, turn the coarse adjustment knob to bring the specimen into focus. After obtaining approximate focus, use the fine adjustment knob to make fine adjustments. (Page 11)



6. Looking through the left eyepiece with your left eye, turn the diopter adjustment ring to focus the specimen. (Page 11)



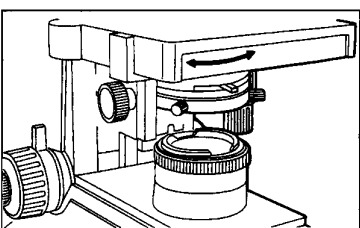
7. Adjust the interpupillary distance of the eyepieces. (Page 11)



8. Center the field iris diaphragm by manipulating the two centering screws on the compensating lens.

9. Engage the objective to be used for observation and adjust the light intensity to the desired level, then readjust the focus.

10. Adjust the field iris diaphragm. (Page 12)



11. Adjust the aperture iris diaphragm. (Page 12)

5 USING THE CONTROLS

5-1 Base

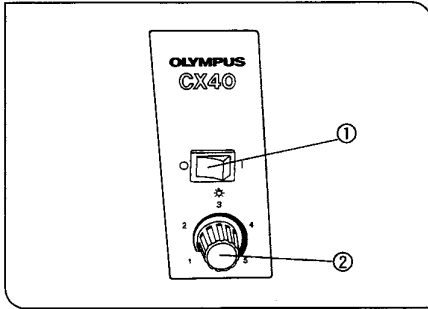


Fig. 13

1 Turning ON the Bulb

(Fig. 13)

1. Switch the main switch ① to "I" (ON).
2. Turning the light intensity knob ② clockwise in the direction of the higher numbers makes the illumination brighter, and vice versa.

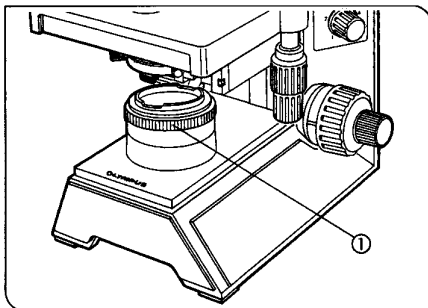


Fig. 14

2 Field Iris Diaphragm

(Fig. 14)

Using the field iris diaphragm dial ①, adjust the diameter of the field iris for objective power to the extent that it just circumscribes the field of view. When the field iris diaphragm is stopped down to circumscribe the field of view, it will exclude extraneous light and improve image contrast within the field of view.

★ When using 100X objective, the field iris diaphragm image will not be visible within the field of view. Accordingly, stop down the diaphragm to its smallest diameter.

3 Dummy Slider

The dummy slider provided with the microscope frame can be used to accommodate the optional transmitted light analyzer (U-ANT).

By preparing a transmitted light polarizer (U-POT) and polarizing light condenser (CH3-CDP), simple polarized light observation becomes possible.

5-2 Stage

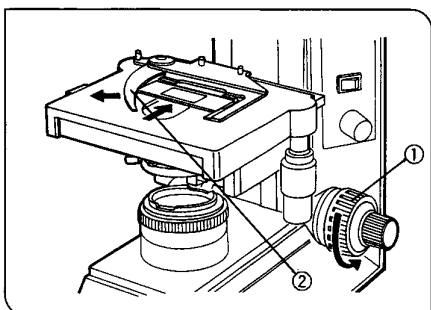


Fig. 15

1 Specimen Placement

(Fig. 15)

- ★ Releasing the curved finger with great force, or suddenly releasing your grip on the curved finger knob while releasing the curved finger, will crack or damage the slide glass. Always place the specimen with great care.

Specimen Holder for Single Slides

1. Turn the coarse adjustment knob ① counterclockwise in the direction of the arrow to lower the stage.
2. Open the spring-loaded curved finger ② on the specimen holder and slide the specimen slide into the specimen holder from the front.
3. After placing the slide as far as it will go, gently release the curved finger ②.

Specimen Holder for 2 Specimen Slides

1. Place the first specimen slide as described in steps 1 and 2 above. Then place the second specimen slide so that it contacts the first specimen slide.
2. Gently release the curved finger ②.

Placing the Specimen Slide with One Hand

Place the specimen slide at the front of the stage, then slide the specimen slide on the stage surface to slowly and gradually open the curved finger in the direction of the arrow. Insert the specimen slide into the specimen holder until it is fully and properly seated in the specimen holder.

● Cover Glass

Use cover glasses of 0.17 mm thickness in combination with objectives marked with the " $\infty/0.17$ " engraving for optimum performance of these objectives.

● Specimen Slide

Specimen slides with a thickness between 0.9 mm and 1.4 mm are recommended for the CX40 microscope. Using thicker specimen slides may result in less than optimum illumination.

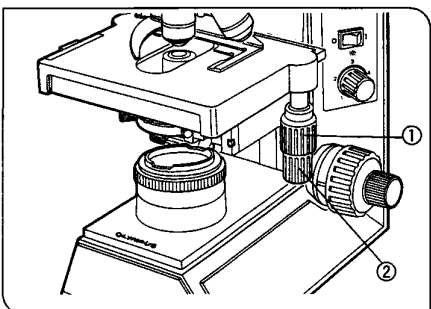


Fig. 16

2 Moving the Specimen

(Fig. 16)

Turning the Y-axis knob ① moves the specimen in the Y-axis direction. Turning the X-axis knob ② moves the specimen in the X-axis direction.

- ★ Never move the specimen by holding the specimen holder or stage directly as this will damage the knob rotation mechanism.
- ★ When further movement of the stage or specimen holder is prevented by the provided limiting mechanisms, the tension of the Y-axis and X-axis knob rotation will increase. At this point, do not rotate the knobs further. Applying excessive force could destroy the limiting mechanism.

5-3 Observation Tube

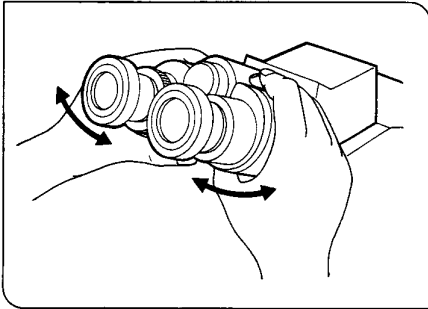


Fig. 17

1 Adjusting the Interpupillary Distance (Fig. 17)

While looking through the eyepieces, adjust the binocular vision until the left and right fields of view coincide completely. The index dot • indicates the interpupillary distance.

- Ⓢ Note your interpupillary distance so that it can be quickly duplicated.

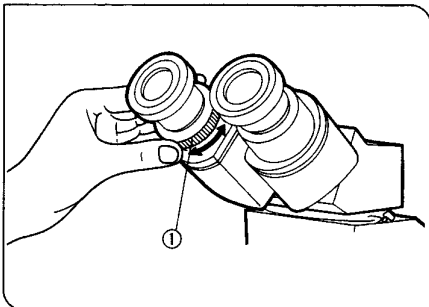


Fig. 18

2 Diopter Adjustment (Fig. 18)

1. Looking through the right eyepiece with your right eye, rotate the coarse and fine adjustment knobs to bring the specimen into focus.
2. Looking through the left eyepiece with your left eye, turn the diopter adjustment ring ① to focus the specimen.

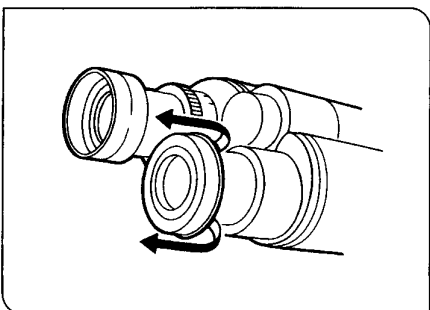


Fig. 19

3 Using the Eye Shades (Fig. 19)

When Wearing Eyeglasses

Use with the eye shades in their normal folded down position. This will prevent the eyeglasses from contacting and scratching the eyepieces.

When Not Wearing Eyeglasses

Extend the folded eye shades in the direction of the arrow to prevent extraneous light from entering between the eyepieces and eyes.

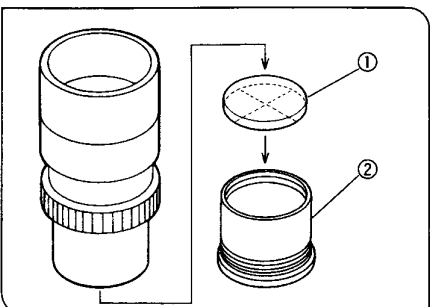


Fig. 20

4 Using Eyepiece Micrometer Discs (Fig. 20)

Eyepiece micrometer discs can be inserted into CWH10X-H (and CWH10X) eyepieces.

Use 24 mm diam. × 1.5 mm micrometer discs. Following Fig. 20, unscrew the micrometer mounting frame ② from the eyepiece and place a micrometer disc ① into the frame. The engraving on the micrometer disc ① should face downward into the micrometer mounting frame ②.

Screw the micrometer mounting frame back into the eyepiece tube.

5-4 Condenser

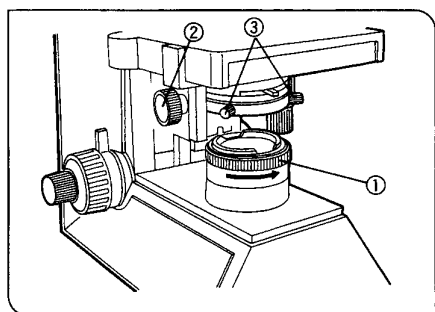


Fig. 21

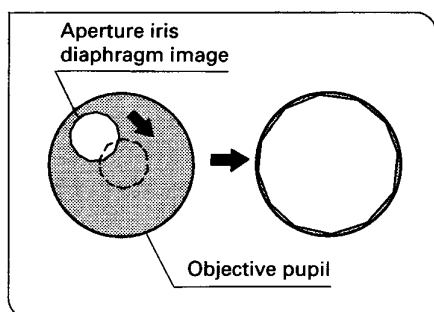


Fig. 22

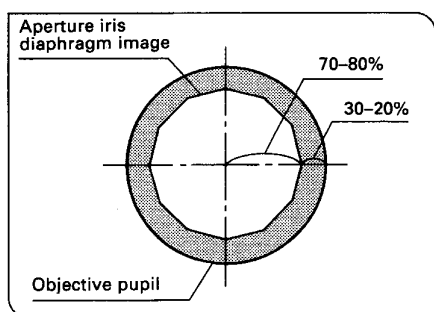


Fig. 23

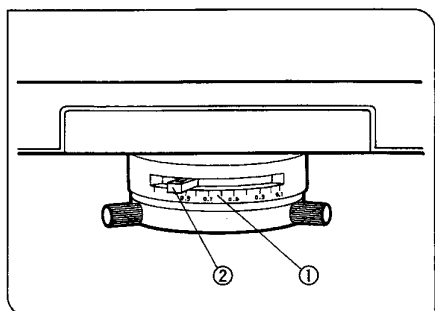


Fig. 24

1 Centering the Field Iris Diaphragm (Figs. 21, 22)

1. With the 10X objective engaged and the specimen brought into focus, turn the field iris diaphragm dial ① counterclockwise to stop down the diaphragm to near its minimum size. (Fig. 21)
2. Turn the condenser height adjustment knob ② to bring the field iris diaphragm image into focus. (Fig. 21)
3. While gradually opening the field iris diaphragm image, rotate the two centering knobs ③ of the compensating lens to adjust so that the field iris diaphragm image is centered in the eyepiece field of view. (Figs. 21, 22)
4. To check centration, open the field iris diaphragm until its image touches the perimeter of the field of view. If the image is not precisely inscribed in the field of view, center again. (Fig. 22)
5. When used for actual observation, open the field iris diaphragm until its image just circumscribes the field of view.

2 Aperture Iris Diaphragm (Figs. 23, 24)

- The aperture iris diaphragm determines the numerical aperture of the illumination system. Matching the numerical aperture ① of the illumination system with that of the objective provides better image resolution and contrast, and also increases the depth of focus.
 - Since the contrast of microscope specimens is ordinarily low, setting the condenser aperture iris diaphragm to 70-80% of the N.A. of the objective in use is usually recommended. When necessary, adjust the ratio by removing the eyepiece and looking into the eyepiece sleeve while adjusting the aperture iris diaphragm lever ② until the image shown in Fig. 23 is seen. (Fig. 24)
- ◎ **Using the Numerical Aperture Scale**
Align the aperture iris diaphragm knob ② with the NA value ① on the scale. The scale value should correspond to the NA number engraved on the objective.
Since the aperture iris diaphragm knob has a certain width, align the line engraved on the knob with the scale indication.
When using 100X oil objective, turn the aperture iris diaphragm knob ② all the way to 0.9 on the scale. (Fig. 24)

5-5 Focusing Adjustment Knobs

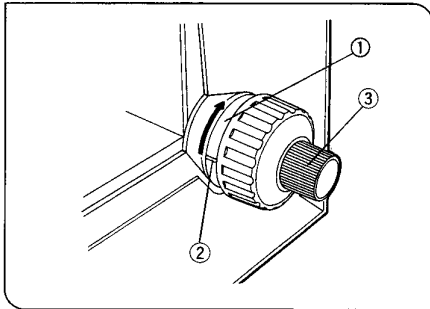


Fig. 25

1 Adjusting the Coarse Adjustment Knob Tension (Fig. 25)

1. The coarse adjustment knob tension is preadjusted for easy use. However, if desired, one can change the tension using the tension adjustment ring ①. Applying a large flat-bladed screwdriver to any of the grooves ② on the circumference of the ring and turning the ring clockwise in the direction of the arrow increases tension, and vice versa.
2. The tension is too low if the stage drops by itself or focus is quickly lost after adjustment with the fine adjustment knob ③. In this case, turn the ring ① in the direction of the arrow to increase tension.

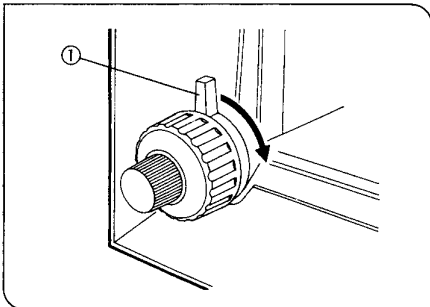


Fig. 26

2 Pre-focusing Lever (Fig. 26)

The pre-focusing lever ensures that the objective does not come in contact with the specimen and simplifies focusing. After focusing on the specimen with the coarse adjustment knob, turn this lever ① clockwise in the direction of the arrow to set an upper limit on coarse adjustment movement.

- ⊙ Focusing with the fine adjustment knob is not affected by this pre-focusing lever. Accordingly, after using the coarse adjustment knob to lower the stage for changing specimens or applying immersion oil (see Section 5-6), refocusing is easily accomplished by rotating the coarse adjustment knob to reach the pre-focusing position, then making fine adjustments with the fine adjustment knob.

★ When not required, leave the pre-focusing lever unlocked.

5-6 Immersion Objectives

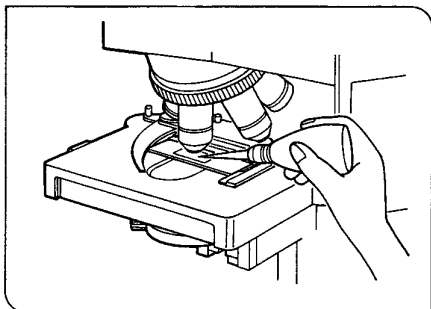


Fig. 27

1 Use of Immersion Objectives (Fig. 27)

1. Focus on the specimen with a low power objective.
 2. Place a drop of immersion oil (provided) onto the specimen at the area to be observed.
 3. Turn the revolving nosepiece to engage the immersion objective, then focus using the fine adjustment knob.
 - ★ Since any bubbles in the oil will impair the image, make sure that the oil is free of bubbles.
 - a. To check for bubbles, remove the eyepiece and fully open the field and aperture iris diaphragms, then look at the exit pupil of the objective inside the observation tube. (The pupil should appear round and bright.)
 - b. To remove bubbles, rock the nosepiece slightly to move the oil immersion objective back and forth a few times.
 - ⊙ If the condenser marking shows a numerical aperture (NA) of 1.0 or more, the number applies only when oil is present between the slide glass and the top surface of the condenser. When oil is not present, the NA is about 0.9.
 4. After use, remove oil from the objective front lens by wiping with gauze slightly moistened with an ether (7 parts) / alcohol (3 parts) mixture.
 - ★ Using too much alcohol can dissolve the lens adhesive.
- ⚠ Caution on Using the Immersion Oil:**
 If the immersion oil comes into contact with your eye or skin, immediately take the following action.
 For eye: Rinse with clean water (for more than 15 min.).
 For skin: Wash with soap and water.
 If the appearance of the eye or skin changes or pain continues, please consult your doctor.

5-7 Photomicrography

- ⊙ Use a trinocular observation tube (U-TR30) for photomicrography. Photomicrography can be performed using the PM-10, the PM-20, or the PM-30 photomicrographic system. Procedures for operating the photomicrographic units are described in their respective instruction manuals. Procedures specific to this microscope are described below.

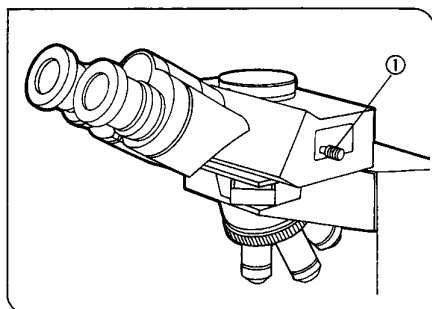


Fig. 28

1 Light Path Selection (Fig. 28)

- Slide the light path selector knob ① to select the desired light path.
- ⊙ The selector knob is ordinarily at the middle position. With dark specimens, push the knob in. If additional light is needed for television or photomicrography, pull the knob out.

Light path selector knob	Symbol	Intensity ratio	Application
Pushed in		100% for binocular eyepieces	Observation of dark specimens
Middle position		20% for binocular eyepieces, 80% for TV/photography	Observation of bright specimens, photography, TV observation
Pulled out		100% for TV/photography	Photography, TV observation

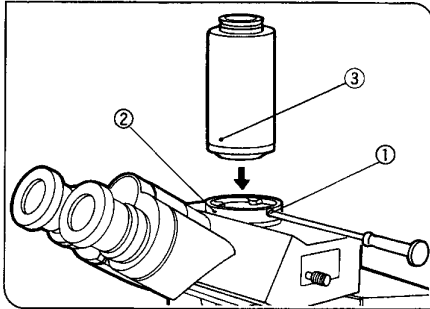


Fig. 29

2 Attaching Straight Photo Tube (U-SPT) (Fig. 29)

1. Using the Allen screwdriver, loosen the clamping screw ① on the trinocular tube photo port.
2. Align the index dot ② with the index dot ③ on the straight photo tube, then mount the straight photo tube on the trinocular tube photo port.
3. Tighten the clamping screw ①.

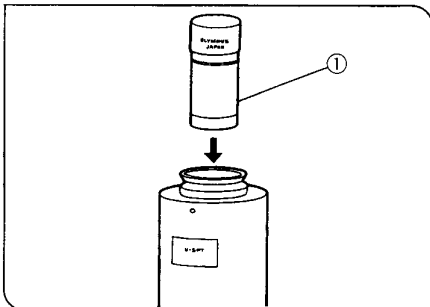


Fig. 30

3 Photo Eyepiece (Fig. 30)

Use the PE photo eyepiece for photomicrography. Insert the photo eyepiece of your choice ① into the straight photo tube mounted on the trinocular observation tube.

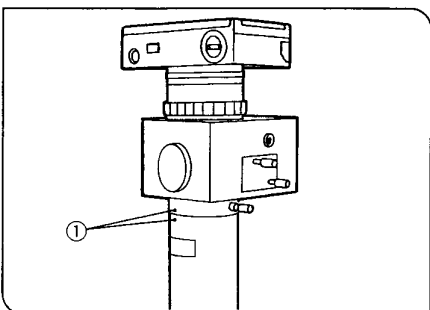


Fig. 31

4 Mounting the Camera Unit (Fig. 31)

Place the camera unit directly over the circular dovetail of the straight photo tube. Make sure the index dots ① on the straight photo tube and the camera unit are aligned, then clamp the unit.

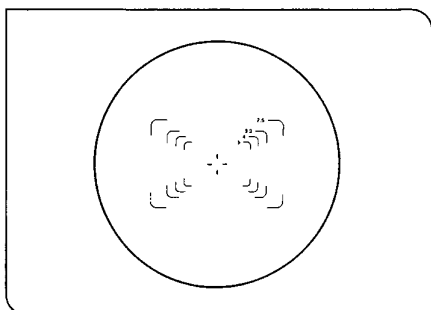


Fig. 32

5 Focus Adjustment

(Fig. 32)

1. Camera focusing is done using the binocular section of the trinocular observation tube.
★ Whenever you remove the focusing telescope from the camera unit, be sure to install the dust cap.
 2. Insert a finder eyepiece into the right eyepiece sleeve.
 3. The finder eyepiece has a built-in focusing lens with four masks and a double crossline, and the focus is practically the same for the focusing lens and the camera film plane. The masks indicate the areas covered, and the numerals next to the masks correspond to the magnification of the photo eyepiece. Different finder eyepieces are available for different cameras. Select the type that is appropriate for the camera being used.
 4. Because of the great depth of focus of 4X and lower objectives, use of the focusing magnifier (U-FT) is recommended for accurate focusing.
- ⊙ When using low power objectives, focusing may be accomplished easier using the focusing telescope on the camera unit rather than the finder eyepiece.

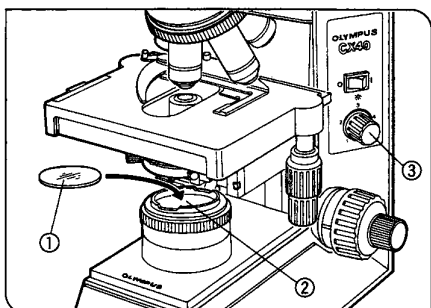


Fig. 33

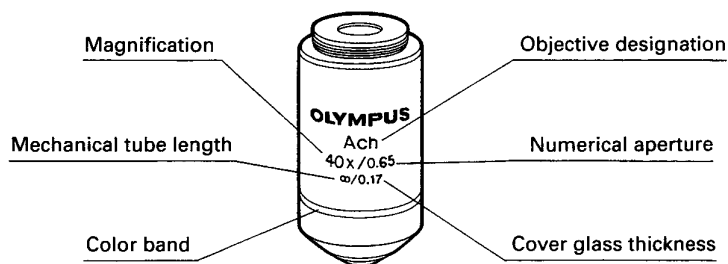
6 Color Temperature Compensation

(Fig. 33)

- ⊙ When using daylight type film in the camera.
- ★ The blue filter is solely designed for visual observation (daylight). If this filter is engaged while taking photographs with daylight film, the color reproduction will be poor.
1. Mount the light balancing filter (45LBD-2N) ① in the filter holder ② of the microscope frame.
 2. Turn the light intensity knob ③ to adjust the brightness of the bulb. At the index the color of the illumination will be close to daylight.
- ⊙ For accurate measurement of the color temperature, it is recommended to use an Olympus color temperature meter (PM-CTR).

Item	Specification	
(1) Optical system	UIS (Universal Infinity System) optical system	
(2) Illumination	Illuminator built-in 6V, 30 W halogen bulb Philips 5761 (Average service time: Approximately 100 hrs. when used as directed) CX40 RF100/LF100: 100/110-120V, 50/60 Hz 0.42A 100-120V area CX40 RF200/LF200: 220/230-240V, 50/60 Hz 0.21A 220-240V area	
(3) Focusing	Stage movement by roller guide (Rack & Pinion) Stroke per rotation: 39.6 mm Full range stroke: 25 mm Upper limit stopper Torque adjustment on coarse focus knob	
(4) Revolving nosepiece	Built-in quintuple nosepiece with inward tilt	
(5) Observation tube	Type	U-BI30 Widefield binocular
		U-TR30 Widefield trinocular observation tube
	Field No.	20
	Tube inclination	30°
	Interpupillary distance adjustment	50 mm–76 mm
Light path selector	None	3 steps: ① Bi 100% ② Bi 20%, Photo 80% ③ TV, Photo 100%
(6) Stage	Size	188 mm × 134 mm
	Movement	76 mm X-direction X 50mm Y-direction
	Specimen holder	Double slide holder
(7) Condenser	Type	Abbe condenser
	N.A.	1.25 (with oil immersion)
	Aperture iris diaphragm	Built-in
(8) Dimensions and weight	W233 × H416 × D377 mm • 9 kg (19.8 lb)	
(9) Operating Environment	<ul style="list-style-type: none"> • Indoor use • Altitude: Max. 2000 m • Ambient temperature: 5° to 40°C (41° to 104°F) • Maximum relative humidity 80% for temperatures up to 31°C (88°F) decreasing linearly through 70% at 34°C (93°F), 60% at 37°C (99°F), to 50% relative humidity at 40°C (104°F) • Main supply voltage fluctuations not to exceed ±10% of the nominal voltage. • Pollution Degree 2 (in accordance with IEC 664) • Installation/Overvoltage Category II (in accordance with IEC 664) 	

7 OPTICAL CHARACTERISTICS



Optical character Objectives	Magnification	N.A.	W.D. (mm)	Cover glass thickness	Resolu- tion (μm)	Eyepiece			Remarks
						CWH10X (FN 20)			
						Total mag.	Depth of focus (μm)	Field of view	
Plan Plan Achromat (FN22)	4X	0.10	22.0	—	3.36	40X	175	5.0	
Ach Achromat (FN22)	10X 40X 100XOI	0.25 0.65 0.60-1.25	6.1 0.45 0.13	— 0.17 —	1.34 0.52 0.27	100X 400X 1000X	28.0 3.04 0.69	2.0 0.5 0.2	Iris

Glossary

Working distance (W.D.):	The distance from the cover glass surface to the nearest point of the objective.
Numerical aperture (N.A.):	The N.A. value represents a performance number which can be compared to the relative aperture (f-number) of a camera lens. The higher N.A., the higher the resolving power.
Resolving power:	The ability to differentiate two points, i.e., the minimum distance by which the objects must be separated in order to be revealed as two separate objects.
Focal depth:	The depth in the image through which the focused image will appear uniformly sharp. As the aperture iris diaphragm is stopped down, the focal depth becomes greater. The greater the N.A. of an objective, the shorter the focal depth.
Field number:	A number that represents the diameter in mm of the image of the field diaphragm that is formed by the lens in front of it.
Field of view diameter:	The actual size of the field of view in millimeters.
Total magnification:	Equals the objective magnification multiplied by the eyepiece magnification.



TROUBLESHOOTING GUIDE

Under certain conditions, performance of this unit may be adversely affected by factors other than defects. If problems occur, please review the following list and take remedial action as appropriate. If you cannot solve the problem after checking the entire list, please contact your local Olympus representative for assistance.

Problem	Cause	Remedy	Page
1. Optical System			
a) Field of view is obscured, or field of view is not evenly illuminated.	Nosepiece is not correctly engaged.	Make sure that the revolving nosepiece clicks properly into place.	7
	The condenser is not correctly attached.	Re-insert the condenser all the way without tilt.	3
	The field iris diaphragm is not properly centered.	Center the field iris diaphragm.	12
	The field iris diaphragm is stopped down too far.	Open the field iris diaphragm.	12
	Dirt/dust on the objective, eyepiece, condenser, and/or light exit glass.	Clean thoroughly.	—
	The dummy slider is not correctly engaged.	Make sure that the dummy slider clicks properly into place.	—
b) Dirt or dust is visible in the field of view.	Dirt/dust on the light exit glass.	Clean thoroughly.	—
	Dirt/dust on the top lens of the condenser.		
	Dirt/dust on the specimen.		
	Dirt/dust on the eyepiece.		
c) The image shows diffraction.	The condenser is lowered too far.	Adjust the condenser height position.	—
	The aperture iris diaphragm is stopped down too far.	Open the aperture iris diaphragm.	12
d) Visibility is poor. •Image is not sharp. •Contrast is poor. •Details are indistinct.	The objective is not correctly engaged in the light path.	Make sure that the revolving nosepiece clicks into place correctly.	4
	Front lens of the objective is dirty.	Clean the objective.	—
	Immersion oil is not being used with an oil immersion objective.	Use immersion oil.	14
	The immersion oil contains bubbles.	Remove bubbles.	14
	Recommended immersion oil not used.	Use the provided immersion oil.	—
	Specimen is dirty. Eyepiece, condenser is dirty.	Clean.	—
e) Part of the image is blurred. The image seems to waver.	The objective is not stopped at the click-stop.	Make sure that the revolving nosepiece clicks into place correctly.	4
	The specimen is not mounted correctly on the stage.	Place the specimen correctly on top of the stage and secure it with the specimen holder.	10
f) The image has a yellow tint.	The blue filter is not engaged.	Engage the blue filter.	4
2. Coarse/Fine Adjustment			
a) The coarse adjustment knob is hard to turn.	The tension adjustment ring is over-tightened.	Loosen the ring.	13
	You are trying to raise the stage with the coarse adjustment knob even though the pre-focusing lever is locked.	Unlock the pre-focusing lever.	13
b) The stage drifts down by itself, or focus is lost during observation.	The tension adjustment ring is too loose.	Tighten the ring.	13
c) Coarse adjustment will not go all the way up.	The pre-focusing lever is keeping the stage down.	Unlock the pre-focusing lever.	13
d) Coarse adjustment will not go all the way down.	The condenser holder is too low.	Raise the condenser holder.	—

Problem	Cause	Remedy	Page
e) The objective makes contact with the specimen before focus is obtained.	The specimen is mounted upside-down.	Mount the specimen correctly.	10
3. Observation Tube			
a) Field of view of one eye does not match that of the other.	The interpupillary distance is incorrect.	Adjust the interpupillary distance.	11
	Incorrect diopter adjustment.	Adjust the diopter.	11
	Different eyepieces are used on the left and right.	Change one eyepiece to match the other so that both sides are the same.	—
	The optical axes are not parallel.	Upon looking into the eyepieces, try looking at the overall field before concentrating on the specimen range. You may also find it helpful to look up and into the distance for a moment before looking back into the microscope.	—
4. Stage			
Image blurs as you move the specimen.	The specimen is not correctly positioned on the stage.	Mount the specimen correctly.	10
5. Objective Change			
Front lens of a high power objective comes into contact with specimen when it is engaged after a low power objective.	Specimen is mounted upside-down.	Mount the specimen correctly.	10
	Cover glass is too thick.	Use a 0.17 mm thick cover glass.	10
6. Electrical System			
a) The bulb does not light.	Bulb is not mounted.	Mount the designated bulb.	3
	The bulb is burned out.	Replace the bulb.	3
	The power cord plug is not connected.	Connect the power cord.	5
b) The bulb is dark.	The voltage selector switch is set incorrectly.	Set the switch to match the local AC mains.	i
c) The bulb burns out almost immediately.	Wrong type of bulb used.	Use the correct bulb type.	3
	The voltage selector switch is set incorrectly.	Set the switch to match the local AC mains.	i

■ PROPER SELECTION OF THE POWER SUPPLY CORD

If no power supply cord is provided, please select the proper power supply cord for the equipment by referring to "Specifications" and "Certified Cord" below:

CAUTION: In case you use a non-approved power supply cord for Olympus products, Olympus can no longer warrant the electrical safety of the equipment.

Specifications

Voltage Rating	125V AC (for 100-120V AC area) or, 250V AC (for 220-240V AC area)
Current Rating	6A minimum
Temperature Rating	60°C minimum
Length	3.05 m maximum
Fittings Configuration	Grounding type attachment plug cap. Opposite terminates in molded-on IEC configuration appliance coupling.

Table 1 Certified Cord

A power supply cord should be certified by one of the agencies listed in Table 1, or comprised of cordage marked with an agency marking per Table 1 or marked per Table 2. The fittings are to be marked with at least one of agencies listed in Table 1. In case you are unable to buy locally in your country the power supply cord which is approved by one of the agencies mentioned in Table 1, please use replacements approved by any other equivalent and authorized agencies in your country.
















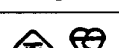



Country	Agency	Certification Mark	Country	Agency	Certification Mark
Argentina	IRAM		Italy	IMQ	
Australia	SAA		Japan	MITI	
Austria	ÖVE		Netherlands	KEMA	
Belgium	CEBEC		Norway	NEMKO	
Canada	CSA		Spain	AEE	
Denmark	DEMKO		Sweden	SEMKO	
Finland	FEI		Switzerland	SEV	
France	UTE		United Kingdom	ASTA BSI	
Germany	VDE		USA	UL	
Ireland	NSAI				

Table 2 HAR Flexible Cord

APPROVAL ORGANIZATIONS AND CORDAGE HARMONIZATION MARKING METHODS

Approval Organization	Printed or Embossed Harmonization Marking (May be located on jacket or insulation of internal wiring)		Alternative Marking Utilizing Black-Red-Yellow Thread (Length of color section in mm)		
			Black	Red	Yellow
Comite Electrotechnique Belge (CEBEC)	CEBEC	<HAR>	10	30	10
Verband Deutscher Elektrotechniker (VDE) e.V. Prüfstelle	<VDE>	<HAR>	30	10	10
Union Technique de l'Electricite' (UTE)	USE	<HAR>	30	10	30
Instituto Italiano del Marchio di Qualita' (IMQ)	IEMMEQU	<HAR>	10	30	50
British Approvals Service for Electric Cables (BASEC)	BASEC	<HAR>	10	10	30
N.V. KEMA	KEMA-KEUR	<HAR>	10	30	30
SEMKO AB Svenska Elektriska Materielkontrollanstalter	SEMKO	<HAR>	10	10	50
Österreichischer Verband für Elektrotechnik (ÖVE)	<ÖVE>	<HAR>	30	10	50
Danmarks Elektriske Materialkontroll (DEMKO)	<DEMKO>	<HAR>	30	10	30
National Standards Authority of Ireland (NSAI)	<NSAI>	<HAR>	30	30	50
Norges Elektriske Materielkontroll (NEMKO)	NEMKO	<HAR>	10	10	70
Asociacion Electrotecnica Y Electronica Espanola (AEE)	<UNED>	<HAR>	30	10	70
Hellenic Organization for Standardization (ELOT)	ELOT	<HAR>	30	30	70
Instituto Portages da Qualidade (IPQ)	np	<HAR>	10	10	90
Schweizerischer Elektro Technischer Verein (SEV)	SEV	<HAR>	10	30	90
Elektriska Inspektoratet	SETI	<HAR>	10	30	90

Underwriters Laboratories Inc. (UL)
Canadian Standards Association (CSA)

SV, SVT, SJ or SJT, 3 X 18AWG
SV, SVT, SJ or SJT, 3 X 18AWG

This device complies with the requirements of both directive 89/336/EEC concerning electromagnetic compatibility and directive 73/23/EEC concerning low voltage. The CE marking indicates compliance with the above directives.

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